
GENETICS

Specificity of *E. coli* K-12 Chromosomal Segment Regulating the Expression of Systems Inhibiting Flac Plasmid Transfer

N. I. Buyanova, V. P. Shchipkov, and A. P. Pekhov

Translated from *Byulleten' Eksperimental'noi Biologii i Meditsiny*, Vol. 124, No. 11, pp. 559-561, November, 1997
Original article submitted December 2, 1996

Specificity of *E. coli* K-12 chromosomal Thr-Leu-segment (genetic locus *tis*) regulating the expression of systems inhibiting Flac plasmid transfer is revealed. The findings point to a complex (polygenous) structure of this locus.

Key Words: *plasmid; chromosomal genes; transfer inhibition system; gene expression*

Chromosome segments of bacterial host cells participate in the regulation of F plasmid and some other F-like plasmid transfer along with plasmid genetic systems (*fin*-systems) [1-4]. Previously, we revealed the role of *E. coli* K-12 Thr-Leu chromosome segment in the expression of *fin* V, an F-like plasmid pAP53 system [2]. Possible effect of chromosome locus, denoted as locus *tis*, on the function of other known systems regulating plasmid transfer is still unclear. In this study we compared the specificity of locus *tis* toward *fin* systems OP, Q, U, V, and W, capable of inhibiting conjugative transfer of Flac plasmid [5,6].

MATERIALS AND METHODS

Reference derepressed (*drd*) Flac plasmid and reference repressed (*rd*) plasmids R100 (*fin* OP), TP108 (*fin* Q), JR66a (*fin* U), R485 (*fin* V), and R455 (*fin* W) from N. Willetts' collection (UK) were used. Plasmid hosts were *E. coli* K-12 cells C600 (Lac, Thr, Leu, and Rif) and AP132 (Lac and Nal). *E. coli* strain Hfr C were donors of the *tis* chromosome locus.

Department of Biology and General Genetics, Russian University of Peoples' Friendship, Moscow

Conjugation crossing-over of bacteria and selection of genetic recombinants and plasmid transconjugates were carried out routinely. For assessing the capacity of *Fin*⁺ plasmids to inhibit the functions of Flac plasmid transfer genes (*Tra*-function), the transfer inhibition index (TII) was calculated as the ratio of the rate of Flac plasmid transfer from single-plasmid donor cells to the rate of the same plasmid transfer from diploplasmid transconjugate cells. Functional activity of "sex" piles whose production is regulated by the Flac plasmid *tra* genes was assessed from the sensitivity of relevant bacterial cells to pile-specific MS2 phage.

RESULTS

Conjugation crossing over of Hfr C (*Tis*⁺) cells with C600 (*Tis*⁻) recipient strain cells was performed to obtain genetic recombinants containing *tis* locus. The resultant Thr⁺Leu⁺ recombinants were used as bacterial hosts containing *drd* Flac plasmid and one reference *rd* plasmid with a known *fin* type. The corresponding single-plasmid transconjugates (recombinant cells containing Flac plasmid alone) were used as control.

Flac plasmid TII for each reference *Fin*⁺ plasmid was estimated from the results of subsequent cross-

TABLE 1. Expression of Plasmid fin Systems in *E. coli* K-12 Cells with Tis⁺ and Tis⁻ Phenotype

Host cell Tis phenotype	Plasmid content of host cell	fin system type of rd plasmid	Flac TII	Tis ⁺ effect value
C600 Tis ⁺	R100+Flac	OP	232-430	7.5-22.5
	TP108+Flac	Q	10-3200	33-3440
	JR66a+Flac	U	1600-31660	10.7-107
	R485+Flac	V	385-406	8.0-10.0
	R455+Flac	W	203-200000	10.0-5263
C600 Tis ⁻	R100+Flac	OP	10-57.6	
	TP108+Flac	Q	0.3-0.93	
	JR66a+Flac	U	149-295	
	R485+Flac	V	35-48	
	R455+Flac	W	20-38	

sings of the resultant diploid and single-plasmid transconjugates with the recipient strain AP132 cells. At least 50 plasmid transconjugates were investigated for identifying the Tis⁺/Tis⁻ phenotype of the resultant recombinants in all cases.

The Thr⁺Leu⁺ recombinants obtained in our experiments are characterized by different TII values, permitting us to divide them into two phenotypic groups (Tis⁺ and Tis⁻, Table 1). For characterization of Tis⁺ effect, its value was determined in each case, i.e., the ratio of Flac TII value derived in study of C600 Tis⁺ cells to similar value for C600 Tis⁻ cells.

Table 1 shows that the value of Tis⁺ effect varies from 7.5 to 5263 and differs appreciably for individual fin systems. The incidence of Tis⁺ effect in Thr⁺Leu⁺ recombinants varied in different fin systems. For fin systems Q and W, Tis⁺ effect was detected in 72 and 68% of all examined Thr⁺Leu⁺ recombinants, respectively. For fin OP, fin U, and fin V, these values varied from 20 to 29%.

A special series of experiments was carried out to assess the specificity of Tis⁺ effect of individual Thr⁺Leu⁺ recombinants, initially detected for R455 rd plasmid (fin W) toward other plasmids carrying other fin systems. The results permit us to identify 3 phenotypic classes of such recombinants. In the first case, Tis⁺ effect was observed only toward fin W system but was completely absent for rd-plasmids with fin

systems OP, Q, and V. In the second case, this effect was observed for fin W and fin OP systems.

Group 3 Thr⁺Leu⁺ recombinants exerted Tis⁺ effect toward fin W group, but fin U system was completely unable to inhibit Flac plasmid transfer, i.e., Flac plasmid TII in this case was even lower than for Tis⁻ cells. The opposite effects of recombinants of this group toward two different fin systems may be due to the genetic structure of *tis* locus.

The results indicate a complex (polygenous) structure of identified chromosome *tis* locus of *E. coli* K-12 cells. The detected differences in the specificity of Tis⁺ effect in different groups of recombinants may be explained by the genetic structure of this locus, which formed as a result of its rearrangement. Further studies are needed to verify this conclusion.

REFERENCES

1. N. I. Buyanova, E. V. Grishina, and A. P. Pekhov, *Byull. Eksp. Biol. Med.*, **116**, No. 10, 424-425 (1993).
2. N. I. Buyanova, V. P. Shchipkov, and A. P. Pekhov, *Ibid.*, No. 9, pp. 306-307.
3. L. Beutin and M. Achtman, *J. Bacteriol.*, **139**, 730-737 (1979).
4. M. Cuozzo and P. M. Silverman, *J. Biol. Chem.*, **261**, 5175-5179 (1986).
5. N. Willetts and R. Skurray, *Annu. Rev. Genet.*, **14**, 41-76 (1980).
6. N. Willetts and R. Skurray, *Escherichia coli and Salmonella typhimurium: Cellular and Molecular Biology*, Vol. 2, Washington (1987), pp. 1110-1133.